- Massanet, G. M. (1974) An. Quím. 70, 74.
- Ando, M., Tajima, K. and Takase, K. (1983) J. Org. Chem. 48, 1210
- Samek, Z., Holub, M., Herout, V. and Sorm, F (1969) Tetrahedron Letters 2931.
- Geissman, T. A. (Ed.) (1962) Chemistry of Flavanoid Compounds, p. 421. Pergamon Press, Oxford.
- 6. González, A. G., Arteaga, J. M. and Bretón, J. L. (1973)
- Phytochemistry 12, 2997.
- 7 Samek, Z. and Holub, M. (1971) Tetrahedron Letters 4775.
- 8. Drozdz, I. B., Holub, M., Zamek, Z., Herout, V. and Sorm, F. (1968) Collect. Czech. Chem. Commun. 33, 1730
- 9 Rustaiyan, A., Nazarians, L and Bohlmann, F (1979) Phytochemistry 18, 879
- González, A. G., Arteaga, J. M. and Bretón, J. L. (1975) *Phytochemistry* 14, 2039.

Phytochemistry, Vol 23, No 9, pp 2072-2074, 1984 Printed in Great Britain.

0031-9422/84 \$3 00 + 0 00 1984 Pergamon Press Ltd

FURANOHELIANGOLIDES AND FLAVONOIDS FROM LOURTEIGIA BALLOTAEFOLIA

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(Received 24 October 1983)

Key Word Index—Lourteigia ballotaefolia; Compositae; Eupatorieae; heliangolides; flavonoids.

Abstract—The aerial parts of Lourteigia ballotaefolia afforded, in addition to three known flavonoids, two furanoheliangolides. The structures and stereochemistries were determined by chemical and spectroscopic means and were established as the 8- and 9-acetylsarracinoyl esters of 1-keto-8 β -9 β -dihydroxygermacra-2,4,11(13)-trien-3, (10 β)-oxo-6 α ,12-olide. The chemotaxonomic situation is discussed briefly.

INTRODUCTION

The genus Lourteigia contains seven species which are distributed in Colombia and Venezuela. This genus is placed in the Gyptis group (tribe Eupatorieae) [1], and to it belongs Eupatorium ballotaefolium H.B.K. which is now named L. ballotaefolia (H.B.K.) K. et R. [2]. No chemical investigations have been reported previously on these plants. We have investigated this species to determine if the chemistry might help in taxonomic revision within this diverse group. The aerial parts of this plant afforded several known flavonoids and two furanoheliangolides which are closely related to those isolated from other genera of this tribe.

RESULTS AND DISCUSSION

The aerial parts of Lourteigia ballotaefolia afforded the flavonoids hispidulin (1) [3], eupafolin (2) [4] and the 3-methyl ether of quercetin (3) [5], as well as two sesquiterpene lactones, 4 and 5, which could only be separated with difficulty. The structure of 4 was identical to conoprasiolide 5'-O-acetate, isolated from Conocliniopsis prasiifolia [6], for which the stereochemistry was given,

*Present address: Departamento de Química Fundamental, Escuela Técnica Superior de Ingenieros Industriales, Universidad Politécnica, Las Palmas de Gran Canaria, Canary Islands, Spain. but the 8β -acyloxy group cannot be assigned with certainty. Acetylation of 4 using pyridine gave a mixture of several compounds. Acetylation using p-toluenesulfonic acid as a catalyst afforded the acetate 6. Catalytic hydrogenation of 4 afforded 7. The C-6 lactone closure was confirmed since the signal at $\delta 5.15$ in the ¹H NMR spectrum of 4 attributed to the C-6 allylic hydrogen of the lactone closure was shifted upfield to $\delta 4.45$ in 7 (Table 1). All the signals were clearly established by decoupling experiments. When 4 was hydrolysed with pyridine it afforded 8. This reaction supports the stereochemistry at C-8 since the hydrolysis under these conditions occurs by an intramolecular catalysis, which is facilitated by the ester and the hydroxyl group in the cis position. The second sesquiterpene lactone was a furanoheliangolide isomer of 4, in which the relative positions of the acetylsarracinate and the hydroxyl group were interchanged. The ¹H NMR spectral data (Table 1) and the obtainment of 8 by hydrolysis with pyridine showed that the structure was 5. The free C-8 hydroxyl group caused a downfield shift of the H-6 signal, again supporting the stereochemistry at these centres. Compounds 4 and 5 were derivatives of atripliciolide, which is the 8-desacyl-9-desoxy derivative of 4 [7].

The sesquiterpene lactones isolated from L. ballotaefolia support the proposed relationship of this genus to the Gyptis group. Obviously more species of the genus Lourteigia need to be investigated but furanoheliangolides have been isolated in the tribe Eupatorieae only from the Gyptis group (Conocliniopsis [6], Trichogonia [8],

$$I = R_1 = R_3 = H; R_2 = OMe$$

2
$$R_1 = H$$
; $R_2 = OMe$; $R_3 = OH$

Table 1. ¹H NMR spectral data of compounds 4-8 (60 MHz, CDCl₃)

	4	5	6	7	8
H-2	5 64 s	5 60 s	5.66 s	5.64 s	5.70 s
H-5	5 98 m	6.00 m	6.00 m	2.58 m	5.90 m
				2 10 m	
I-6	5.15 m	5.75 m	5.40 m	4.45 m	5.20 m
H-7	3 62 m	3.45 m	3.65 m	3 10 m	3.50 m
I-8	5.15 m	4.15 m	5.40 m	5.34 m	4.80 m
1-9	4.18 m	5.28 d	5.32 d	3.95 m	4.10 m
I-13	6.35 d	6.35 d	6.40 d	1.40 d	6.25 d
I-13'	5 73 d	5.68`d	5.85 d		5.60 m
I-14	1.67 s	1.46 s	1.50 s	1.67 s	1.67 s
I-15	2.07 d	2 03 d	2 07 d	1 38 d	205 d
I-3'	7.06 q	7.32 q	7 10 q	7.10 q	
I-4'	1 95 d	2.00 d	1.95 d	1 95 d	
I-5' ₁	4.92 d	5.20 d	4.95 d	4.90 d	
I-5'2	4.67 d	4.83 d	4.65 d	4.65 d	
OAc	2.03 s	2.03 s	2.04 s	2.04 s	
			2.10 s		
Ή	3 62 m	3.45 m			

Bejaranoa [9] and Trichogoniopsis [10]) with the exceptions from Disynaphia halimifolia [11] and Isocarpha atriplicifolia [7].

EXPERIMENTAL

Mps are uncorr. UV were recorded in EtOH. ¹H NMR spectra were recorded at 60 MHz using TMS as internal standard. The aerial parts of the air-dried plant material (3.5 kg) (voucher J. Triana 2 Herbario Fac. de Farmacia, collected in Mérida, Venezuela) was extracted with EtOH at room temp. The resulting extracts were coned and treated with lead acetate soln; the suspension was filtered and extracted with EtOAc. The extract (55 g) was first separated by CC (silica gel) and further by repeated TLC (silica gel). The flavonoids were separated by recrystallization and identified by mps, UV, ¹H NMR spectra and comparison of their acetyl derivatives with published data. 65 mg 1, 114 mg 2, 28 mg 3, 460 mg 4 (EtOAc-petrol, 1:2) and 48 mg 5 (EtOAc-petrol, 1:2) were obtained.

9β-Hydroxyatripliciolide-8-O-(5'-acetoxysarracinate) Colourless gum, $[\alpha]_D^{25} - 82.1^\circ$ (CHCl₃; c 2.1). IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3500 (OH), 1765 (γ-lactone), 1725, 1650 (C=CCO₂R), 1690, 1590 (O=C-C=C-OR); UV $\lambda_{\text{max}}^{\text{EIOH}}$ nm: 210, 265; MS m/z (rel. int): 372 [M - HOAc]⁺ (2), 274 [M - RCO₂H]⁺ (5), 141 [MeCH=C(CH₂OAc)CO]⁺ (61), 99 [141 - ketene]⁺ (20). To 80 mg 4 in 1 ml Ac₂O, 40 mg p-toluenesulfonic acid was added. After 12 hr, CC (EtOAc-petrol, 2.1) afforded 60 mg 6, colourless gum; IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1760 (broad), 1715 (C=O), 1650, 1595 (C=C); UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm: 265. Compound 4 (142 mg) was hydrogenated in the presence of palladium on C (5%) (8 hr). CC (C₆H₆-EtOAc, 3:2) afforded 40 mg 7, colourless crystals, mp 147-148° (EtOAc-petrol). $[\alpha]_D^{25} + 23.5^\circ$ (CHCl₃; c 0.79). IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3470 (OH), 1770 (y-lactone), 1725, 1690 (C=O), 1590 (C=C); UV $\lambda_{\text{max}}^{\text{EiOH}}$ nm: 262; MS m/z (rel. int.): 436.171 [M]⁺ (17) ($C_{22}H_{28}O_9$), 394 [M – ketene]⁺ (5), 376 [M – HOAc]⁺ (9), 141 [MeCH=C(CH₂OAc)CO]⁺ (50). A soln of 4 (120 mg) in C₅H₅N (3 ml) was left to stand for 12 hr at room temp. Work-up in the usual manner and CC (C₆H₆-EtOAc, 3:1) afforded 32 mg 8, colourless crystals, mp 198-200° (EtOAc). $[\alpha]_{\rm D}^{25} - 152^{\circ}$ (CHCl₃; c 0.321). IR $\nu_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 3400 (OH), 1770 (γ-lactone), 1695 (C=O), 1650, 1590 (C=C). MS m/z (rel. int.) 292

 $[M]^+$ (0.2), 274 $[M - H_2O]^+$ (1.2).

9 β -(5'-Acetoxysarracinoyloxy)-atripliciolide (5). Colourless gum $[\alpha]_D^{25}$ -63.6° (CHCl₃; c 0.245). IR $\nu_{max}^{CHCl_3}$ cm⁻¹: 3490 (OH), 1760 (γ -lactone), 1715, 1650 (C=CC $_2$ R), 1700, 1598 (O=C-C=C-OR); UV λ_{max}^{EtOH} nm: 212, 264; MS m/z (rel. int.): 432.142 [M]+ (4) (C₂₂H₂₄O₉), 372 [M-HOAc]+ (2), 274 [M-RCO₂H]+ (4), 245 [274-CHO]+ (2), 232 [274-C₂H₂O]+ (7), 141 [MeCH=C(CH₂OAc)CO]+ (31), 99 [141-ketene]+ (22). 23 mg 5 treated with C₅H₅N as above afforded 8 (TLC and IR spectrum).

Acknowledgements—We thank Prof. L. Ruiz Terán (*) from Facultad de Farmacia, Universidad de Los Andes, for the classification of the plant and the CDCH-ULA for financial support. We are grateful to Dr. F. Bohlmann (Institute for Organic Chemistry, Technical University, Berlin) for supplying the IR and ¹H NMR spectra of conoprasiolide 5'-O-acetate.

REFERENCES

- Robinson, H. and King, R. M. (1977) The Biology and Chemistry of the Compositae (Heywood, V. H., Harborne, J. B. and Turner, B. L., eds.), p. 455. Academic Press, London.
- 2. King, R. M. and Robinson, H. (1972) Phytologia 23, 307.
- 3. Herz, W. and Sumi, Y. (1964) J Org. Chem. 29, 3438.
- Kupchan, S. M., Sigel, C. W., Hemingway, R. J., Knok, J. R. and Udayamurthy, M. S. (1969) Tetrahedron 25, 1603.
- 5 Breton, J. L., Gonzalez, A. G and Rincones, M. (1969) Anal. Quim. 63, 297.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1980) Phytochemistry 19, 1547.
- Bohlmann, F., Mahanta, P. K., Natu, A. A., King, R. M. and Robinson, H. (1978) Phytochemistry 17, 471.
- Bohlmann, F., Zdero, C., Pickard, J., Robinson, H. and King, R. M. (1981) Phytochemistry 20, 1323.
- Bohlmann, F., Abraham, W. R, Robinson, H. and King, R. M. (1981) Phytochemistry 20, 1639.
- Bohlmann, F, Zdero, C., King, R. M. and Robinson, H. (1982) Phytochemistry 21, 2035
- Bohlmann, F., Dhar, A. K., Jakupovic, J., King, R. M. and Robinson, H (1981) Phytochemistry 20, 1077.